

Microbes, Nutrients, and Twitter: What We Have Learned from a WRRI Student Grant Danielle Winter, Bryan Maxwell, François Birgand



Treatment Side

Control Side

What Are Floating Islands? Hydroponic plant growth systems that

- Improve water quality
- · Provide aquatic and terrestrial habitat
- Regulate water temperature
- · Enhance aesthetics

Supply organic matter and dissolved O₂

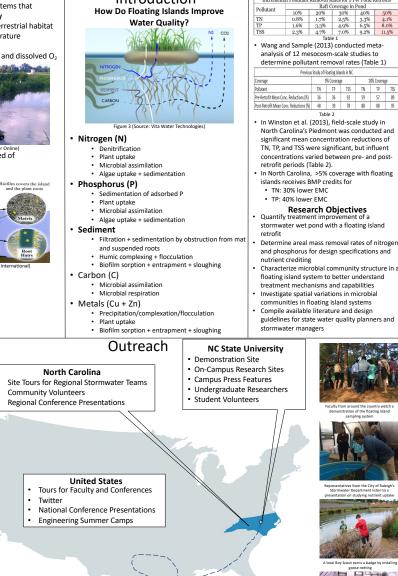


Floating islands are comprised of

- Native wetland plants
- Porous floating media

- Applications include
 - Stormwater
 - Swine lagoons
 - Aquaculture waste
 - Algal control
 - Erosion control

lents rest after a long day o g to set-up the field study



Introduction





guidelines for state water quality planners and

How much coverage is enough?

Incremental Pollutant Removal Rates for FTW Pond Retrofits

Methods Mesocosm-Scale Study

Vegetated floating islands and unvegetated floating islands were placed in individual mesocosms in a greenhouse. Mesocosms were filled with enriched water from a local stormwater/irrigation pond (Figures 4 & 5).

Turbidity and concentrations of NO₂- and DOC were measured with a field spectrophotometer in each mesocosm at a high temporal resolution

One mesocosm trial was conducted in Fall 2017 and two additional trials with be conducted in Summer 2018



Microbial Community Analyses Solution pipetted from floating island mats, root wash from composite

root sample (3 species), and surface sediments were collected weekly. Grab samples and sediment samples were also collected weekly for chemical analyses.

FAME (fatty acid methyl ester) analysis (Agilent 780B Gas Chromatography system) was utilized to detect fatty acids from living

and dead cells and identify organism type based on certain fatty acids that are specific to cell membranes of certain organisms (Table 3).

Fatty Acid	Organism Type	Fatty Acid	Organism Type
i13:0 3OH	Bacteria (Gram -)	cy19:0	Bacteria (Gram -)
i15:0	Bacteria (Gram +)	20:0	Bacteria (cyanobacteria)
a15:0	Bacteria (Gram +)	16:0	Fungi
i17:0	Bacteria (Gram +)	16:1w5	Fungi (AMF)
a17:0	Bacteria (Gram +)	18:1œ9	Fungi
cy17:0	Bacteria (Gram -)	18:2@6	Fungi
i17:0 3OH	Bacteria (Gram -)	18:306	Fungi
17:1@6	Bacteria	10Me 16:0	Actinomycetes
18:1 0 5	Bacteria	10Me 17:0	Actinomycetes
i19:0	Bacteria	10Me 18:0	Actinomycetes
a19:0	Bacteria	20:406	Protozoa

Preliminary Results Field-Scale Study

0.5 E Contro FTW Date 0.5 1.0 1.5 Contro
FTW Date Figure 12 Contro FTW

In October, NO3⁻ concentrations were slightly lower in the Tx section between events (Figure 11) After October, NO₃ concentrations were not consistently higher in either section (Figure 12) Reductions in NO₃⁻ concentration on Tx side do not reflect NC BMP credit, but monitoring has only occurred during dormant season so far (lower plant uptake and microbial activity) Turbidity is consistently higher in the control section (Figure 13)

Field-Scale Study Field site is located in stormwater wet pond (9000 ft²) on NC State's west campus near Wolf Village apartments Pond was split with impermeable harrier into Tx and control sections (~2500 ft²/side) (Figure 6) Tx side has 14 4'X8' islands that were grown for >18 months prior to

Methods

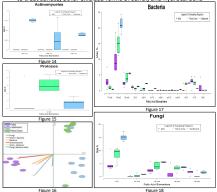


installation in a baffle formation of 3

- Weirs evenly divide flow at inlet of Tx and control sections Riser outlets for each section are split (Figure 7)
- 1 automated sampler collects flow-weighted samples during events at the inlet of the pond
- Each split outlet has an automated sampler that collects flowweighted samples during events (Figures 8) Field spectrophotometer is used for continuous, multi-point sampling during and between events in Tx and control sections
- (Figure 9)
- Sampling at 10-minute resolution for NO₃ and turbidity
- 2 points near the outlet in each section (Figure 10) Grab samples that are collected immediately after and between events and flow-weighted sampling from inlet and outlets during events undergo
- · Laboratory analysis for calibration of spectrophotometer for NO3 and TSS (based on turbidity)
- · Additional laboratory analysis for TN and TP

Preliminary Results **Microbial Community Analyses** Presence of a distinct cluster for each sar

- unique community structures (Figure 16)
- Sediment appears to provide the most conducive environment for biomarkers of bacteria (except for 17:1w6 and i19:0), fungi, and actinomycetes (Figures 14, 17, 18)
- Root environment provides a unique source of nutrition and/o habitat that supports protozoa communities (Figure 15) .
- Root zone and mat samples tended to exhibit more variability than sediment samples · Dynamic communities and environments
 - Shorter establishment period
- Fungal biomarkers were observed in all locations and in substantial concentrations, indicating potential for floating islands to degrade a
- variety of organic compounds, such as PAHs, EDCs, and pesticides Correlations between shifts in sediment biomarkers and water
- quality parameters suggest significant exchange at sediment-water interface
- Sulfate-reducing bacteria (17:1ω6) were observed in all three locations, especially in mats, suggesting potential for floating islands to treat sulfate, other oxidized forms of sulfur, and hydrocarbons





Campus Presence

